© UNAAB 2010

Journal of Natural Sciences, Engineering and

A PRELIMINARY SURVEY ON THE PARASITES OF FREE RANGE CHICKEN IN ABEOKUTA, OGUN STATE, NIGERIA

^{1*}U.F. EKPO, ¹A.A. OGBOOYE, ¹A.S. OLUWOLE AND ²M. TAKEET

¹Department of Biological Sciences, University of Agriculture, Abeokuta, Nigeria ²Department of Veterinary Microbiology and Parasitology, University of Agriculture, Abeokuta, Nigeria.

*Corresponding author: ekpouf:unaab.edu.ng

Tel: +234802755689

ABSTRACT

A preliminary study was carried out to assess the parasitic burden of domesticated and free range chicken in Abeokuta, Ogun State, Nigeria. A total of twenty birds where purchased from five popular markets in Abeokuta namely, Itoku, Kuto, Lafenwa and Osiele and examined for presence of parasites. The skin and feathers of the birds were also combed to check for ectoparasites. Afterward, the birds were slaughtered and the intact gastro-intestinal tract were then separated and opened to check for endoparasites. All parasites found were collected into petri-dishes and examined under microscope for identification. Two species of ectoparasites *Menacanthus stramineus* with a prevalence of 90.0% and *Lipeurus caponis* with a prevalence of 60.0% were recovered. Three species of endoparasites found consisted of two nematodes (*Ascaridia galli* and *Heterakis gallinarum*) and one cestode (*Raillietina echinobothridia*). *Ascaridia galli* and *Raillietina echinobothridia* had the highest prevalence of 80.0% followed by *Ascaridia galli* and *Heterakis gallinarum* each with 60.0% prevalence. The results show that free-range chickens in Abeokuta carry high parasitic burden which could be a big constraint to their productivity and commercial value.

Key words: Endoparasite, Ectoparasites, Free range, chickens, Abeokuta, Nigeria

INTRODUCTION

The poultry industry has an important position in the provision of animal protein (meat and egg) to man and plays a vital role in the national economy as a source of revenue. Poultry is one of the most intensively reared of the domesticated species and one of the most developed and profitable animal production enterprises (Obiora, 1992).

In Nigeria, backyard poultry represents about 60.0% of the 140 million poultry population, thus, the most important form of poultry production (Ikpi and Akinwunmi, 1981). It is estimated that this provide 12kg of poultry needs per inhabitants per year whereas cattle provides 5.3kg (Forsido, 1986). This means that by comparison, poultry meat is more available to the people than beef.

Parasitism has been identified as one of the major factors that threatens village freerange chicken production (Adene and Dipeolu, 1975). Reports have shown that mortality due to parasitic diseases is higher

J. Nat. Sci. Engr. Tech. 2010, 9(2):123-130

¹U.F. EKPO, ¹A.A. OGBOOYE, ¹A.S. OLUWOLE AND ²M. TAKEET

than those attributed to Newcastle disease, and other viral infection of poultries. Common poultry parasites range from lice, mites, fleas, ticks, and helminths to gnats and coccidia (Nnadi and George, 2010). Parasitic infection or their concurrent infections has been known to result in immunosuppression, especially in response to vaccines against some poultry diseases et al., 2003). Helminths (Horning infestations are known to cause interference with host metabolism resulting in poor feed utilization and reduced growth rate as well as size and age at maturity (Nnadi et al., 2007). Studies in other countries had shown that the prevalence of parasitic infestations in village chicken flocks is close to 100.0%, and in most cases individual birds' harbour more than one parasite type (Permin et al., 1997). It is believed that, understanding of parasitic diseases of birds will help in devising the measures to improve health and utility of these birds (Msoffe et al., 2010). The present study is therefore a survey to determine preliminary the prevalence of ecto and endo parasites of local chicken in Abeokuta.

MATERIALS AND METHODS Study animals

Twenty adult free range chickens were bought from Osiele, Itoku, Lafenwa and Kuto markets. Five chickens were bought from each market. They were of indigenous breed of both sexes of which 40.0% of the birds were brown, 30.0% were black, 20.0% were grey and 10.0% were white.

Examination of chicken for ectooparasites

The body of the chickens, including the head, cloacal, bronchial, ventral and femoral areas were combed for ectoparasites. The parasites were removed with camel hair brush, transferred and preserved in 10% formol saline.

Examination of chicken for endoparasites

The chickens were then slaughtered and the intact gastro-intestinal tracts were then separated into oesophagus, crop, proventiculus, gizzard, intestine and caecum. Each part was opened and its content was emptied separately into labelled beakers. Contents were washed into petri-dishes and examined under a stereo microscope. Parasites found were counted and stored in bottles containing formalin.

Identification of parasites

Identification of parasites was carried out at the Department of Veterinary Microbiology and Parasitology, College of Veterinary Medicine, University of Agriculture, Ogun State. Nigeria. All parasites were identified under light microscope with 40x - 100x magnification, using identification keys of Walker *et al.*, (2003).

Data Analysis

The data collected from the study were entered and analysed with SPSS version 11. Chi square analysis was used to compare differences in prevalence of parasites.

RESULTS

Of the 20 chickens examined, nine (45%) were males while 11(55%) were females. The prevalence of ecto and endo parasite in the chicken population by sex is shown in Table1. Statistical analysis at (p>0.05) shows no significant difference in infection status among the sexes for both ecto and endoparasite. All (100%) the chickens examined were infected with endoparasites. Three species of endoparasites were identified and these consist of two nematodes (Ascaridia galli and Heterakis

A PRELIMINARY SURVEY ON THE PARASITES OF FREE RANGE CHICKEN IN...

gallinarum) one cestode (Raillietina echinobothridia). There was no trematode infection. The prevalence of the species of endoparasite significantly is different (p=0.025) among parasite species. The most common infection observed in the chicken population is with the cestode; Raillietina echinobothridia 16 (80%) and the least common were nematodes Ascaridia galli 12 (60%) and Heterakis gallinarum 12(60%).

Out of two nematodes identified one was found in the small intestine and the other in the caecum and the only cestode parasite was found in the small intestine (Table 2). There was no parasite in the gizzard and trachea. The prevalence of nematode parasites was significantly higher than that of cestode parasite (p=0.0000).

Also all the chickens examined were infected with one or two species of ectoparasites. These were lice. Infestation with *Menacanthus stramineus* was significantly higher (p=0.028) than infestation with *Lipeurus caponis* with prevalence of 18 (90%) and 12 (60%) respectively. Also the number of *Menacanthus stramineus* (71) collected was significantly (p=0.0000) more than *Lipeurus caponis* (21).

The pattern of infection with ecto and endo parasite is shown in Table 3.Mixed infections with endoparasites were found in 75% of the chickens, while 25% had single infections. Among the mixed infections, 25% had triple infection 50% had double infection. Mixed infections with ectoparasites occurred as double infections in 50% of the chickens while 50% had single infections. There was no significant difference in the pattern of

Parasites		MALE		FEMALE				
Endoparasite	Number examined	Number infected	% infected	Number examined	Number infected	% infected	P value	
Ascaridia galli	9	3	33.33	11	9	81.82	0.081	
Raillietina echinobothridia	9	7	77.78	11	9	81.82	0.736	
Heterakis gallinarum Ectoparasite	9	6	66.67	11	6	54.55	0.927	
Menacanthus stramineus	9	8	88.89	11	10	90.91		
Lipeurus Caponis	9	5	55.56	11	7	63.64	0.927	

Table 1: Prevalence of infection by sex of free range chicken examined in Abeokuta

1U F EKPO	1 A A	OGBOOYE	1AS	OLUWOLE	AND	² M. TAKEET
0.1° EKI 0.1°	, _1.,_1.	OODOOTE,	11.0.	OLUWOLL	π	WI. 1711XL2L21

Parasite	Location of parasite in the	Parasite count	Number of Infected	% prevalence of parasite in	% prevalence of parasite spp
Endoparasite	body		chickens	chicken population	
Ascaridia galli	Small intestine	51	12	60	41.80
Raillietina echinobothridia	Small intestine	40	16	80	32.79
Heterakis gallinarum	Caecum	31	12	60	25.41
P value				0.3012	0.025
Ectoparasite					
Menacanthus stramineus	Skin	71	18	90	72.45
Lipeurus caponis	Feather	27	12	60	27.55
P value				0.028	0.000

Table 2: Prevalence of infection by location in the chicken's body in Abeokuta n (Endoparasite) =122, n (Ectoparasite)=98

Table 3: Pattern of infection with Ecto and Endo parasites of free range chicken in Abeokuta

Endoparasite	Male	Female	Total (%)
Single Infection	3	2	5 (25)
Double infection	5	5	10 (50)
Triple infection	1	4	5 (25)
P value			0.413
Ectoparasite			
Single Infection	5	5	10(45)
Double infection	4	6	1050
P value			0.653

ecto parasite in chicken population sampled is shown in Table 4. There was no significant difference in mean parasite burden for all parasite species between sexes of chicken.

The mean parasite burden with endo and All the chicken irrespective of type of plumage were infected with both ecto and endo parasite (Table 5). A comparison between endo and ecto - parasites collected from the chicken population revealed that infection with endoparasite is significantly higher (p=0.02) than infestation with

Parasites MALE							
Endoparasite	Mean	SD	Range	Mean	SD	Range	P value
Ascaridia galli	1.2222	1.9221	3-5	3.6363	2.2482	2-6	0.1841
Raillietina echinobothridia	1.6667	1.1180	1-3	2.2727	1.4894	1-4	0.324
Heterakis gallinarum Ectoparasite	1.6667	1.8028	1-5	1.4545	1.6949	1-5	0.7891
Menacanthus stramineus	3.7778	1.7159	3-6	3.3636	1.6293	2-6	0.5863
Lipeurus caponis	1.3333	1.3229	2-3	1.3636	1.2060	1-3	0.9578

Table 4: Mean Parasite, standard deviation and range of parasites in Male and Female chickens

Table 5: Prevalence of ecto and endo parasites by Plumage of free range chicken in Abeokuta

	Ectoparasite			Endoparasite		
Plumage	Number examined	Number infected	% infected	Number examined	Number infected	% infected
Black	6	6	100	6	6	100
White	2	2	100	2	2	100
Brown	8	8	100	8	8	100
Grey	4	4	100	4	4	100

DISCUSSION

The result of this study showed the presence of parasitic infestations among free range chickens in Abeokuta. The high prevalence of endoparasitism (100%) in the present study might be a result of open range conditions that facilitate infection of birds. Local chickens satisfy their nutrient requirement by roaming from place to place and they usually seek their food in the

superficial layers of the soil which is often contaminated with parasites eggs and larva of all kinds, including various insects or earthworm that serve as paratenic or intermediate hosts for endoparasites that infest poultry (Muhairwa *et al.*, 2007 Puttalakshmamma *et al.*, 2008). The prevalence reported here is similar to those of (Hove *et al.*, 2002; Sam-Wobo and Mafiana, 2003) who reported 100 and 98%

J. Nat. Sci. Engr. Tech. 2010, 9(2):123-130

¹U.F. EKPO, ¹A.A.OGBOOYE, ¹A.S.OLUWOLE AND ²M. TAKEET

respectively of endoparasite infection. The multiple presence of endoparasitic infections shows the serious risk of infection in local chickens kept in Africa (Kimani, et al., 1999; Ashenafi and Eshutu, 2004). Studies have also shown that almost 100% of local chickens are infected with helminths parasites (Permin et al., 1997; Horning et al., 2003). The total numbers of intestinal helminths observed in this study are lower than those reported by Sam-Wobo and Mafiana (2003) in Abeokuta, Nigeria and Permin et al., (2002) in Zimbabwe and Ashenafi and Eshutu (2004) in Ethiopia, this may be due to the small number of chicken examined in this study. The three species of intestinal helminths identified in this study are most prevalent intestinal helminths of birds reported (Horning et al., 2003). The percentage prevalence of Raillietina echinobothridia observed from this study is higher than the observations of Sam-Wobo and Mafiana (2003) and Ashenafi and Eshutu (2004) with prevalence of 35.9 and 65.3% respectively. The percentage prevalence of Ascaridia galli, 60% was lower compared to similar observations by Permin et al., (2002) and Sam-Wobo and Mafiana (2003) which had 69 and 73.4% respectively but was higher than the studies done by Kimani et al. (1999) and Ashenafi and Eshutu (2004) with prevalence of 10.03 and 55.3% respectively. The prevalence of Heterakis gallinarum was 60% which was found to be higher than those done by Kimani et al. (1999) and Ashenafi and Eshutu (2004) with a prevalence of 21.33 and 32.6% respectively. However in the present study none of the birds harboured trematode parasites, (this might be due to non accessibility of infected snails. (Puttalakshmamma et al., 2008).

All the chickens examined for ectoparasites were infected (100%). This result is similar to those of Hove et al. (2002), Njunga (2003) and Salam et al. (2006), which had prevalence of 100 and 97.69% respectively. The two species of ectoparasite identified from the study were lice out of which Menacanthus stramineus was more prevalent. Earlier reports of similar studies have indicated lice infestation as the most outstanding infestation among ectoparasite of chickens (Benbrook, 1965 and Fabiyi, 1988) Menacanthus stramineus had a prevalence of 90% which was higher than the observations of Mungube et al. (2005) in Kenya and Belihu (2009) in Ethiopia with prevalence of 79.4 and 65.5% respectively but was similar to the reports of Hove et al. (2002) with a prevalence of 88%. Lipeurus caponis had a prevalence of 60% which was higher than the percentages reported by (Salam et al. 2006) in India with, (Lalitha et al., 2008) and (Eneanya et al., 2008) in South East Nigeria with a prevalence of 39.66, 29.1 and 41.61% respectively. There was no observations of infestation with fleas in this study this is similar to the findings of (Adene and Dipeolu, 1975). However, Nnadozie and George (1996) reported fleas as the dominant ectoparasites in domestic chickens in their survey of blood and ecto-parasites of domestic fowls in Ibadan, Western Nigeria while Salfina (1990) reported fleas as the least occurring of ectoparasites of birds.

The result of this study shows that there was no significant difference in infection status for sex and plumage of the chickens. It means that chickens of all sexes and colours are vulnerable to parasite infection.

CONCLUSION

This study clearly indicated that free range chickens in Abeokuta, Nigeria carry high

burden of parasitic infections. This is associated with their indiscriminate scavenging behaviour. Despite the absence of any veterinary care for free range village birds, their contribution to supply of eggs, meat for household consumption and income generation cannot be mistreated. It is therefore; necessary that control and preventive measures with better management system should be provided for keepers of local chickens so as to boost the poultry production sector of agriculture.

ACKNOWLEDGEMENTS

The authors wish to thank the Technologist of Department of Veterinary Microbiology and Parasitology, College of Veterinary Medicine, University of Agriculture, Abeokuta for his assistance in the identification of the parasites.

REFERENCES

Adene, D.F., Dipeolu, O.O. 1975. Survey of blood and ectoparasites of domestic fowls in Ibadan, Western State of Nigeria. *Bulletin Animal. Health and Production in Africa*, 23: 333-

Ashenafi, H., Eshutu, Y. 2004. Study on Gastro-intestinal Helminthes of local chickens in Central Ethiopia. *Revue de Medecine Veterinaire*. 155(10):504-507.

Belihu, K. 2009. Prevalence of ectoparasites in backyard local chickens in three agroecologic zones of East Shoa, Ethiopia. *Revue de Medecine Veterinaire*. 160 (11):537-541.

Benbrook, E.A. 1965. External parasites of poultry. In Biester, H. E and Schwart L.H. 482 edn. Diseases of poultry. 5th edn .Iowa State University Press, USA. Pp 925-962.

Eneanya, C.I., Amagba I.C., Ikpeze, O.O. 2008. Ectoparasitic fauna of poultry farms in Awka, Anambra, Nigeria. *International Journal of Zoonoses*, 13(2):93-97.

Fabiyi, J.P. 1988. Lice infestation in domestic chicken in Nigeria. *Bulletin Of Animal of Health. Prod. Africa*, 36:390-394.

Forsido, T. 1986. Studies on the meat production potential of some local strains of chickens in Ethiopia. *Justus Liebig University, Giessen, Germany, PhD Thesis.*

Horning G, Rasmussen S., Permin A., M. Bisgaard, 2003. Investigation on the influence of Helminths parasites on vaccination of chicken s against Newcastle diseases Virus under village conditions. *Tropical Animal health and Production*, 35: 415-424

Hove, T., Permin, A., Esmann, J.B., Hoj, C.H., Mukaratirwa, S. 2002. Ecto, endo and haemoparasites in free range chickens in the Goromonzi District in Zimbabwe. *Preventive Veterinary Medicine*, 54(3):213-224.

Ikpi, A, Akiniwumi J. 1981. The future of the poultry industry in Nigeria, *World Poultry Sciences Journal*, 37(1): 39-43.

Kimani, R.N., Irungu, L.W., Kisia. S.M. 1999. Intestinal parasites of chickens in Kenya. *Kenyan Journal of Agricultural Science*, 13:67-74.

Lalitha, J., Rani, N., Basith, S. 2008. Prevalence of ectoparasites of chicken reared under different systems of management. *Journal of Veterinary Parasitology*, 22(2):45-50.

Msoffe P.L.M., Muhairwa A.P., Chiwanga, G.H., Kassuku A.A. 2010. A

J. Nat. Sci. Engr. Tech. 2010, 9(2):123-130

study of ecto and endo-parasites of domestic pigeons in Morogoro Municipality, Tanzania. *African Journal of Agricultural Research*, 5(3):264-267Available o n l i n e a t <u>h t t p : / /</u> www.academicjournals.org/AJAR

Muhairwa, A.P., Msoffe, P.L., Ramadhani, S., Mollel, E.L., Mtambo, M.M.A., Kassuku, A.A 2007. Prevalence of gastro-intestinal helminthes in free-range ducks in Morogoro Municipality, Tanzania *Livestock Research for Rural Development* 19(4):

Mungube, E.O., Bauni, S.M., Ten hagen, B.A., Wamae, L.W., Nzioka, S.M., Muhammed, L., Nginyi, J.M. 2005. Prevalence of parasites of the local scavenging chickens in a selected semi-arid zone of Eastern Kenya. *Tropical Animal Health and Production*, 40(2):101-109.

Njunga, G.R. 2003. Ecto and Haemoparasites of chickens in Malawi. *Preventive Veterinary Medicine*, 54(3):213-224.

Nnadi, P.A Kamalu, T.N., Onah, D.N. 2007 "The effect of dietary protein supplementation on the pathophysiology of Haemonchus contortus infection in West African Dwarf goats," *Veterinary Parasitology*, 148: 256–261

Nnadi, P.A., George, S.O. 2010 A cross sectional survey on parasites of chickens in selected villages in the sub-humid zones of South-eastern Nigeria. *Journal of Parasitology Research* In press.

Obiora, F.C. 1992. A guide to poultry Production in the Tropics. 1st edn. Acena Publishers. Pp. 59-61;. 381, 382

Permin, A., Magwisha, H., Kassuku, A.A., Nansen, P., Bisgaard, A., Frandsen, F., Gibons, L. 1997. A cross sectional study of helminthes in rural scavenging poultry in Tanzania in relation to season and climate. *Journal of Helminthology*, 71: 233-240.

Permin A., Esmann J.B., Hoj C.H., Hove T. Mukaratirwa S. 2002 Ecto-, endo- and haemoparasites in free-range chickens in the Goromonzi District in Zimbabwe. *Preventive Veterinary Medicine*,45: 213-224.

Puttalakshmamma G.C., Ananda, K.J., Prathiush, P.R. Mamatha, G.S., Suguna Rao. 2008 Prevalence of Gastrointestinal parasites of Poultry in and around Bangalore. *Veterinary World*, 1(7): 201-202

Salfina, A.M. 1990.Study of Gastrointestinal helminthes of scavenging chickens in Kalimantana, Indonesia. *Journal of Kansas Entomology*, 29(2):63-79.

Salam, S.T., Mir M.S., Khan, A.R. 2010. The prevalence and pathology of *Raillietina cesticillus in* indigenous chicken (Gallus gallus domesticus) in the temperate Himalayan region of Kashmir - short communication *Veterinarski Arhiv.*, 80(2): 323-328.

Sam-Wobo, S.O., Mafiana, C.F. 2003. Prevalence and Identification of Helminth Parasites of local chickens of Abeokuta, Nigeria. *Asset Series*, 2(2):141-147.

Walker, A.R., Bouattour, A., Camicas, J.L., Estrada- Pena, A., Horak, I.G., Latif, A., Pegram, R.G., Preston, P.M. 2003. Ticks of domestic animals in Africa, A guide to identification of species. *Bioscience*

(Manuscript received: 2nd August, 2010; accepted: 1st May, 2011).